

Rickettsiales and Chlamydiales

Rickettsiales

- Minute obligate intracellular Gram negative bacteria
- Rods or coccobacilli
- Variable size 0.3-0.6 micron
- Non-motile
- Aerobic , divide by binary fission
- Cell wall similar to Gram negative bacteria but in anaplasmatacea have cells bound by two-layer membrane

General features

- Stain poorly with basic dyes
- Stain well with Giemsa and other Romanowsky stains
- Most require living cells for propagation
- Cultured commonly in fertile eggs, or in tissue cultures
- *Haemobartonella felis* not yet propagated in vitro

Key points

- Minute, non-motile Gram-negative bacteria Obligate intracellular pathogens, replicating only in cells
- Demonstrated in blood smears by Romanowsky stains
- Host specificity and tropism for particular cell types evident
- * Extracellular survival brief for most members apart from ***Coxiella burnetii***
- Cause systemic diseases, mainly arthropod borne, in humans and animals
- ***Rickettsiaceae***
 - --cell walls often contain peptidoglycan
 - --cultured in specific cell lines or in fertile eggs
 - -tropism for vascular endothelium or leukocytes
- ***m Anaplasmataceae***
 - -lack cell walls, possess cell membranes
 - -have not been cultured ***in vitro***
 - --tropism for erythrocytes

General features

- They are parasites of arthropods, replicating in cells of gut
- Passed transovarially or transtadially in ticks and mites
- Labile outside the host except *Coxiella brunetti* (resistant due to spores)

Table 111. Genera of the Rickettsiales of veterinary importance.

Family and tribe	Genus	Cells parasitised in host
<i>Rickettsiaceae</i>		
Tribe Rickettsieae	<i>Rickettsia</i>	Vascular endothelial cells
	<i>Coxiella</i>	In vacuoles of cells of reticuloendothelial system
Tribe Ehrlichieae	<i>Ehrlichia</i>	Circulating leukocytes
	<i>Cowdria</i>	Vascular endothelial cells
	<i>Neorickettsia</i>	Reticular cells of lymphoid tissue
<i>Anaplasmataceae</i>		
	<i>Anaplasma</i>	Within erythrocytes
	<i>Aegyptianella</i>	Within erythrocytes of birds
	<i>Haemobartonella</i>	On, or in, erythrocytes
	<i>Eperythrozoon</i>	On erythrocytes

Various Rickettsiales of veterinary significance

Agent	Main host(s) and distribution	Transmission (vector)	Disease and cells parasitised
<i>Rickettsia rickettsii</i>	Humans, dogs. Western hemisphere, especially Eastern USA	Endemic in ticks (<i>Dermacentor</i> spp.). Bites of infected ticks	Rocky Mountain spotted fever in man. Tick fever in dogs. Parasitises vascular endothelial cells
<i>Coxiella burnetii</i>	Humans, cattle and small ruminants. Other animals can be a reservoir. Worldwide	Agent in milk, birth fluids of ruminants and dust. Persists in ticks	Q fever in man: aerosol inhalation, ingestion, tick bites or laboratory accidents. Occasional abortions, weak offspring or infertility in ruminants; usually a tick-borne infection
<i>Ehrlichia canis</i>	Dogs. Americas, Asia, Africa, Caribbean and Mediterranean	Brown dog tick (<i>Rhipicephalus sanguineus</i>)	Canine ehrlichiosis (tropical canine pancytopenia). Agent in lymphocytes, monocytes and, rarely, in neutrophils
<i>E. equi</i>	Horses. USA only	Vector unknown, ticks suspected	Equine ehrlichiosis. Granulocytes and vascular endothelial cells affected
<i>E. ondiri</i>	Cattle. East Africa	<i>Amblyomma</i> ticks	Bovine petechial fever. Granulocytes and monocytes parasitised
<i>E. (Cytoecetes) phagocytophila</i>	Cattle, sheep and wild ruminants. UK, Ireland and Scandinavia	<i>Ixodes ricinus</i>	Tick-borne fever. Pregnant animals may abort. Present in granulocytes and monocytes
<i>E. platys</i>	Dogs. Similar distribution to <i>E. canis</i>	Vector not known	Canine infectious cyclic thrombocytopenia. Usually subclinical. Platelets affected
<i>E. risticii</i>	Horses. USA and Europe	Vector unknown. Seasonal, late spring to early autumn	Potomac horse fever (equine monocytic ehrlichiosis). Present in monocytes. Intestinal involvement
<i>Cowdria ruminantium</i>	Domestic and wild ruminants. Africa, Caribbean	<i>Amblyomma</i> ticks	Heartwater. Reticular cells, neutrophils, and vascular endothelial cells affected
<i>Neorickettsia helminthoeca</i>	Dogs, coyotes, foxes, bears and ferrets. Pacific coast of USA	Ingestion of salmon containing the agent	Salmon poisoning. Agent present in salmon within the metacercariae of the <i>Nanophyetus salmincola</i> fluke. Parasitises reticuloendothelial cells of the lymphoid system, including macrophages
Elokomin fluke fever agent	As above	As above	Salmon fever (milder form). This agent occurs with <i>N. helminthoeca</i> or separately. Reticuloendothelial cells of the lymphoid system

Agent	Main host(s) and distribution	Transmission (vector)	Disease and cells parasitised
<i>Anaplasma marginale</i>	Cattle and other ruminants. Tropical and sub-tropical regions	Ticks and other arthropods and veterinary instruments	Anaplasmosis (gall sickness). Inside erythrocytes with a marginal distribution
<i>A. centrale</i>	As above	As above	Mild or subclinical disease. Can be used to protect animals against the more virulent <i>A. marginale</i> . Predominantly central in erythrocytes
<i>A. ovis</i>	Sheep, goats and deer	As above	Infrequent and mild disease. Predominantly marginal in erythrocytes
<i>Aegyptianella pullorum</i>	Poultry. South Africa, South-east Asia and Mediterranean	Ticks such as <i>Argas persicus</i>	Aegyptianellosis. Agent in erythrocytes, leukocytes and mononuclear cells. <i>A. persicus</i> can also transmit <i>Borrelia anserina</i>
<i>Haemobartonnella felis</i>	Cats. Worldwide	Possibly via cat fights and biting arthropods. Intrauterine infection reported	Feline infectious anaemia. Present on or in erythrocytes
<i>Eperythrozoon ovis</i>	Sheep. Africa, USA, Australia and Europe	Biting arthropods	An infrequent disease in lambs with anaemia and failure to gain weight. Present on red cells
<i>E.suis</i>	Pigs. USA	Biting arthropods and instruments	Porcine eperythrozoonosis. Usually subclinical but occasionally icterohaemia, embryonic death and abortion. Present on erythrocytes

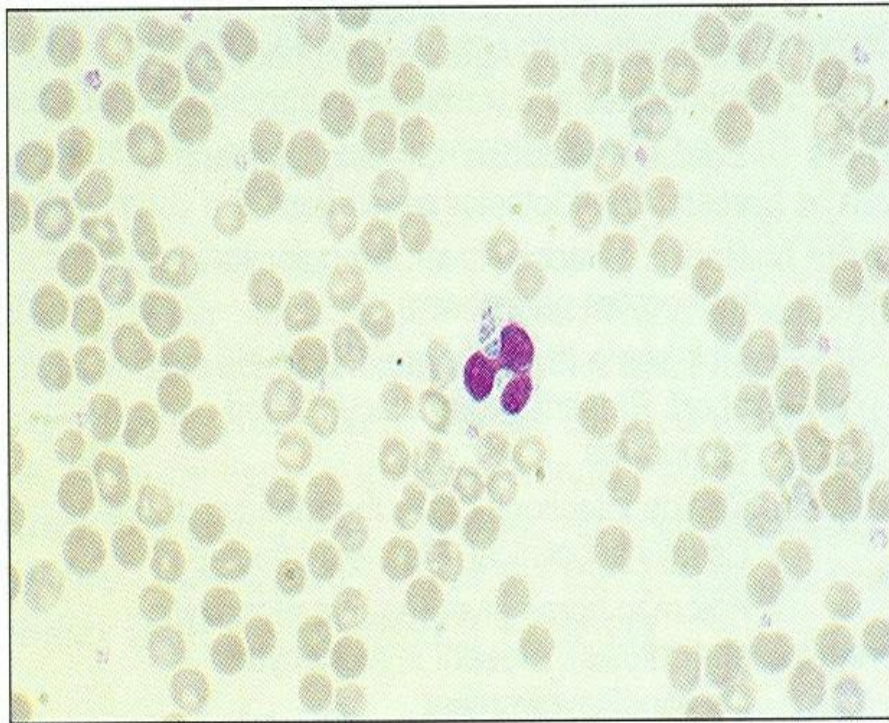
Lab diagnosis

- **Specimen**

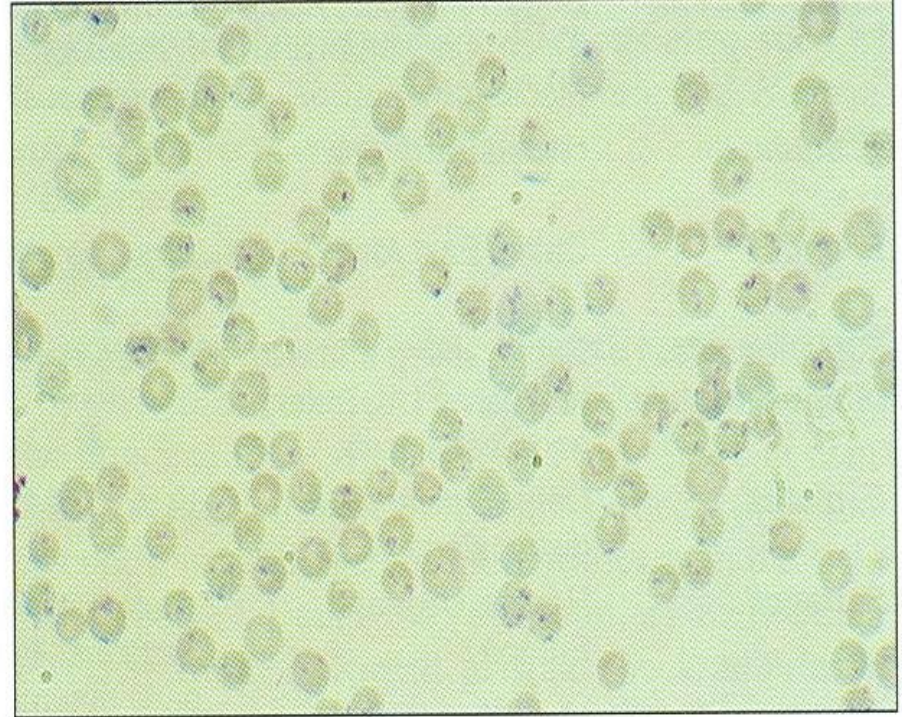
- Vary with disease, may include
- 1. unclotted blood
- 2. affected tissue like brain (heart water)
- 3. paired serum

- **Direct microscopy**

- Poorly stain with Gram stain
- Giemsa and other Romanowsky stains like Giemenez, Machiavello and leishman used
- FAT staining for blood and tissue smears

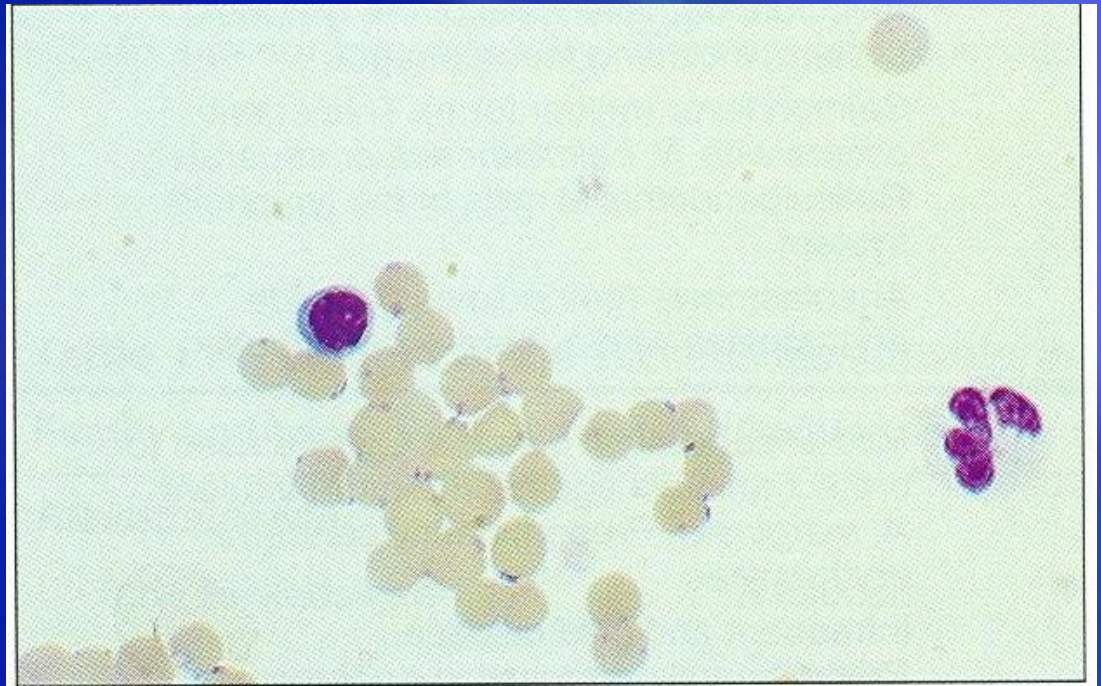


371 Leishman-stained blood smear from a sheep showing a neutrophil containing *Ehrlichia (Cytoecetes) phagocytophila*. (×1000)

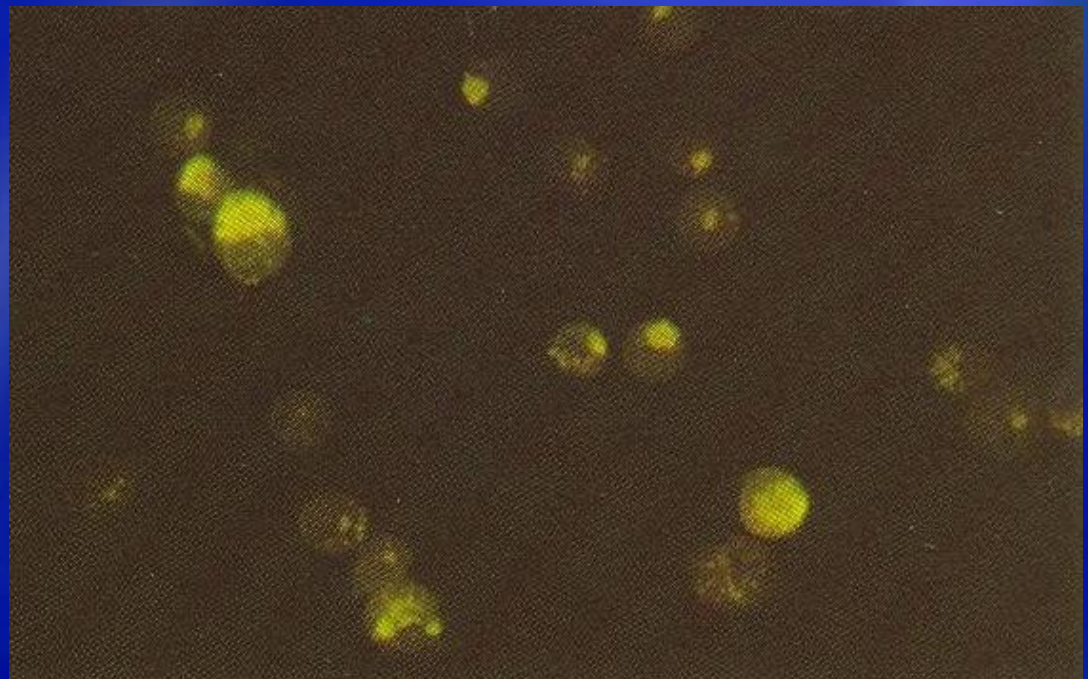


372 Leishman-stained blood smear from a sheep with *Eperythrozoon ovis* parasitising the red cells. (×1000)

**Leishman-stained
blood smear from a cat
with *Haemobatonella
felis***



***Ehrlichia risticii* in
mouse macrophages
(FAT, X1000)**



Isolation and cultivation

- Often difficult
- *Rochalimaea* will grow on inert media but other Rickettsiales require living cells for replication
- Yolk sac of chick embryos, cell cultures or lab animals
- **Identification**
- Based on animal species, clinical signs and demonstration of organism in stained blood or tissue smears and by specific serological tests

- Serological tests
- Q fever
- Weil felix reaction

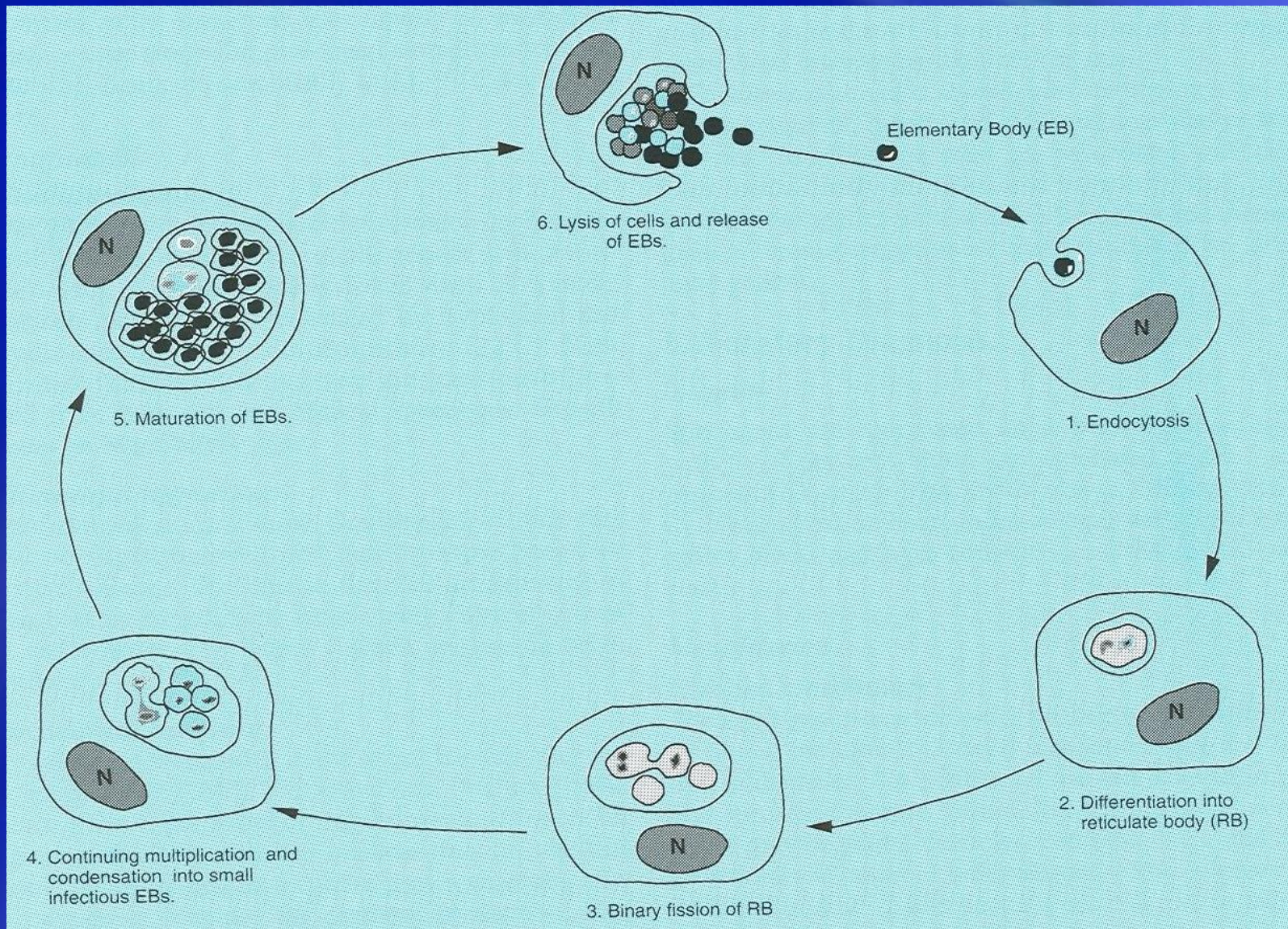
Chlamydiales

- Order Chlamydiales consist of one family Chlamydiaceae with one genus Chlamydia
- Posses cell wall similar to Gram negative but lack muramic acid
- Obligate intracellular parasites with requirement of high energy compounds like ATP
- Also called ***energy parasites***
- Developmental cycle consist alternating, morphologically distinct, infectious and reproductive forms

- *Elementary body is small (200-300 nm), infectious and represents extracellular form of organism*
- EB enters cell by endocytosis and differentiates into larger non-infectious but metabolically active reticulate body inside an expanding vacuole
- RB multiply by Binary fission producing further RBs
- At present only two species are recognised
 - *C. trachomatis*
 - *C. psittaci*

Key points

- Spherical intracellular bacteria with unique developmental cycle
- Appropriate staining procedures include the modified Ziehl-Neelsen and Giemsa methods
- Unable to synthesize ATP and replicate only in living cells
- Cell walls lack peptidoglycan but contain genus-specific lipopolysaccharide
- Species vary in virulence for particular hosts; some strains are associated with specific diseases in domestic animals
- Produce respiratory, enteric, plural and reproductive tract diseases in animals and humans



Representation of the life cycle of chlamydiae (N = Nucleus).

Table 110. Clinical conditions associated with chlamydial infections.

<i>Chlamydia</i> species	Host species	Clinical condition
<i>C. trachomatis</i>	Human	<ul style="list-style-type: none"> • Ocular disease <ul style="list-style-type: none"> a) Trachoma b) Inclusion conjunctivitis of neonates • Genital tract infections <ul style="list-style-type: none"> a) Non-gonococcal urethritis b) Salpingitis c) Cervicitis d) Epididymitis • Respiratory disease of infants • Proctitis • Lymphogranuloma venereum • Arthritis
<i>C. psittaci</i> (TWAR strains) (<i>C. pneumoniae</i>)	Human	<ul style="list-style-type: none"> • Pneumonia
<i>C. psittaci</i>	Human (zoonosis)	<ul style="list-style-type: none"> • Psittacosis (ornithosis) • Abortion • Conjunctivitis
<i>C. psittaci</i>	Mammalian	<ul style="list-style-type: none"> • Intestinal infection and diarrhoea • Gastritis • Pneumonia • Abortion, e.g. enzootic abortion of ewes, chlamydial abortion of cows • Genital infections • Mastitis • Polyarthritis/polyserositis • Encephalomyelitis, e.g. sporadic bovine encephalomyelitis • Hepatitis • Conjunctivitis, e.g. feline pneumonitis
<i>C. psittaci</i>	Avian (psittacosis/ornithosis)	<ul style="list-style-type: none"> • Pneumonia and air sacculitis • Pericarditis • Encephalitis • Conjunctivitis • Intestinal infection and diarrhoea

Lab diagnosis

- **SPECIMEN**

- **Abortion-**

- stained smear from affected cotyledons/vaginal swabs/wet surface of aborted fetus
- Pieces of cotyledon/ foetal liver/ lung

- **Polyarthriti-**

- aspirated synovial fluid

- **Conjunctiviti-**

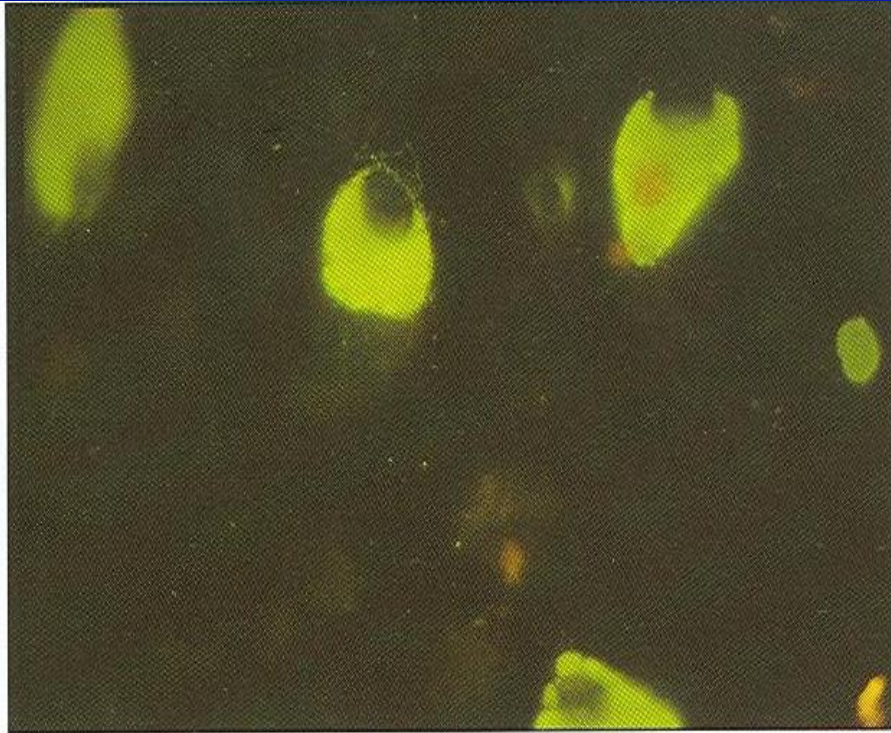
- conjunctival swab

- **Systemic infection-**

- samples of lung, liver and spleen

Isolation and cultivation

- Mouse inoculation
- Inoculation of embryonated eggs(6-7days) via yolk sac route
- Cell culture-McCoy, baby hamster kidney-21, L929 and Vero
- **Serology**
 - CFT
 - EIISA
 - Indirect FAT



368 *C. psittaci* (EAE isolate) inclusions in McCoy cells.
(Direct FA technique, ×400)



369 *C. psittaci* (EAE) inclusions in McCoy cells.
(Giemsa stain, ×400)

THE END